



Prenatal phthalate exposure in relation to placental corticotropin releasing hormone (pCRH) in the CANDLE cohort

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ARTICLE INFO

Handling Editor: Martí Nadal

Keywords:

Phthalates
Endocrine disrupting chemicals
Pregnancy complications
Corticotropin releasing hormone
Placenta

ABSTRACT

Context: Phthalates may disrupt maternal-fetal-placental endocrine pathways, affecting pregnancy outcomes and child development. Placental corticotropin releasing hormone (pCRH) is critical for healthy pregnancy and child development, but understudied as a target of endocrine disruption.

Objective: To examine phthalate metabolite concentrations (as mixtures and individually) in relation to pCRH.

Design: Secondary data analysis from a prospective cohort study.

Setting: Prenatal clinics in Tennessee, USA.

Patients: 1018 pregnant women (61.4% non-Hispanic Black, 32% non-Hispanic White, 6.6% other) participated in the CANDLE study and provided data. Inclusion criteria included: low-medical-risk singleton pregnancy, age 16–40, and gestational weeks 16–29.

Intervention: None.

Main outcome measures: Plasma pCRH at two visits (mean gestational ages 23.0 and 31.8 weeks) and change in pCRH between visits (Δ pCRH).

Results: In weighted quantile sums (WQS) regression models, phthalate mixtures were associated with higher pCRH at Visit 1 ($\beta = 0.07$, 95 %CI: 0.02, 0.11) but lower pCRH at Visit 2 ($\beta = -0.08$, 95 %CI: -0.14 , -0.02). In stratified analyses, among women with gestational diabetes ($n = 59$), phthalate mixtures were associated with lower pCRH at Visit 1 ($\beta = -0.17$, 95 %CI: -0.35 , 0.0006) and Visit 2 ($\beta = -0.35$, 95 %CI: -0.50 , -0.19), as well as greater Δ pCRH ($\beta = 0.16$, 95 %CI: 0.07, 0.25). Among women with gestational hypertension ($n = 102$), phthalate mixtures were associated with higher pCRH at Visit 1 ($\beta = 0.20$, 95 %CI: 0.03, 0.36) and Visit 2 ($\beta = 0.42$; 95 %CI: 0.19, 0.64) and lower Δ pCRH ($\beta = -0.17$, 95 %CI: -0.29 , -0.06). Significant interactions between individual phthalate metabolites and pregnancy complications were observed.

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<https://doi.org/10.1016/j.envint.2022.107078>

Received 26 October 2021; Received in revised form 20 December 2021; Accepted 2 January 2022

Available online 7 January 2022

0160-4120/© 2022 The Authors.

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Conclusions: Phthalates may impact placental CRH secretion, with differing effects across pregnancy. Differences in results between women with and without gestational diabetes and gestational hypertension suggest a need for further research examining whether women with pregnancy complications may be more vulnerable to endocrine-disrupting effects of phthalates.

1. Introduction

Phthalates are synthetic chemicals that are commonly used in consumer products, resulting in widespread human exposure through ingestion, inhalation, and dermal routes (Wang et al., 2019). Many studies including the National Health and Nutrition Examination Survey (NHANES), have demonstrated that nearly 100% of individuals sampled- including pregnant women- have measurable levels of one or more phthalate metabolites in their urine (Swan et al., 2015; Woodruff et al., 2011; Ferguson et al., 2019; Arbuckle et al., 2014). Urinary phthalate metabolite measurement is the preferred method of phthalate exposure assessment in humans and concentrations reflect recent exposures, given phthalates' short-half life in the body (several hours) (Koch et al., 2006). Extensive evidence in animal models indicates that phthalates are endocrine disruptors and reproductive toxicants and, increasingly, epidemiological studies suggest similar developmental and reproductive impacts in humans (Dorman et al., 2018; Martínez-Razo et al., 2021). For example, a number of studies have examined the potential impact of phthalates on pregnancy complications (including gestational diabetes (James-Todd et al., 2016; Shaffer et al., 2019; Zukin et al., 2021; Fisher et al., 2018) and hypertensive disorders of pregnancy (Werner et al., 2015; Cantonwine et al., 2016; Philips et al., 2019) as well as preterm birth (Ferguson et al., 2014; Ferguson et al., 2014; Ferguson et al., 2019; Gao et al., 2019; Hu et al., 2020; Meeker et al., 2009; Zhong et al., 2021) and subsequent offspring developmental outcomes (Swan et al., 2015; Day et al., 2021; Harley et al., 2019; Shoaff et al., 2017). More subtle, subclinical impacts on pregnancy physiology have also been noted including disruption of placental hormone production and activity (reviewed in (Warner et al., 2021).

Largely overlooked in the literature on placental impacts of phthalate exposure is placental corticotropin releasing hormone (pCRH), a 41-amino acid neuropeptide which is produced by the placenta and rises exponentially across gestation (reviewed in (Thomson, 2013). The identical molecule is produced by the paraventricular nucleus of the hypothalamus in both pregnant and non-pregnant individuals and plays a critical regulatory role in the hypothalamic–pituitary–adrenal (HPA) axis, the body's major stress pathway (Taylor and Fishman, 1988). During pregnancy, maternal pCRH concentrations are 10,000 times higher than the CRH levels observed in non-pregnant individuals, thus pCRH measured in circulation during pregnancy is assumed to be almost exclusively of placental origin. In animal studies, pCRH regulates pathways involved in myometrial contraction, promoting labor (Smith et al., 2002; Tyson et al., 2009); and in epidemiological studies higher and/or more steeply increasing pCRH in mid-pregnancy is linked to increased odds of subsequent preterm birth (Hobel et al., 1999; Makrigiannakis et al., 2007; Sandman et al., 2006; Ruiz et al., 2016; Wadhwa et al., 2004). pCRH has been additionally linked to maternal hypertensive disorders of pregnancy (Harville et al., 2009; Laatikainen, 1991), maternal postpartum depression (Yim et al., 2009; Glynn and Sandman, 2014), and offspring developmental outcomes (Howland et al., 2016; Sandman, 2018). Consistent with its role in the HPA axis, some studies have found pCRH to be responsive to socioenvironmental factors, such as experiences of trauma. For example, prior work has demonstrated that pCRH production was higher and rose more steeply among women who reported childhood trauma (Moog et al., 2016; Steine et al., 2020).

Despite increasing interest in psychosocial predictors of pCRH, until recently, other types of stressors, such as environmental exposures, have been largely ignored. In the Puerto Rican PROTECT cohort, Cathey et al (Cathey et al., 2019) reported that concentrations of several maternal

urinary phthalates (including monocarboxyisononyl phthalate [MCNP], mono-(3-carboxypropyl) phthalate [MCP], mono-2-ethyl 5-carboxypentyl phthalate [MECPP], mono (2-ethyl-5-hydroxyhexyl) phthalate [MEHHP], and mono-(2-ethyl-5-oxohexyl) phthalate [MEOHP], but not mono(2-ethylhexyl)phthalate [MEHP]) were inversely associated with pCRH concentrations (Cathey et al., 2019). By contrast, in an *in vitro* model, MEHP treatment increased pCRH protein and mRNA levels and promoted pro-labor gene pathways (Wang et al., 2016). To our knowledge, these associations have not been examined in other epidemiological studies. Our objective in this study was to build upon this limited literature to examine maternal phthalate concentrations in relation to pCRH in mid and late pregnancy, additionally examining potential moderators including pregnancy complications, fetal sex, and maternal history of childhood trauma. Pregnancy complications and history of childhood trauma were selected for consideration due to their associations with pCRH in prior work (Harville et al., 2009; Moog et al., 2016; Steine et al., 2020; Laatikainen et al., 1991), whereas fetal sex was considered in light of the extensive evidence that phthalates (like many endocrine disruptors) can have differential effects on male and female fetuses (and by extension their placentas) (Warner et al., 2021).

2. Methods

2.1. Study population and overview of study activities

The Conditions Affecting Neurocognitive Development and Learning in Early Childhood (CANDLE) study recruited pregnant women receiving care at selected prenatal clinics in Shelby County, Tennessee from 2006 to 2011 and methods have been described elsewhere in detail (Sontag-Padilla et al., 2015). Inclusion criteria for participation included: low-medical-risk singleton pregnancy, 16–40 years of age, weeks 16–29 of pregnancy, and intending to deliver at a participating medical center. Low-medical-risk was defined as lacking major medical conditions including (but not limited to) chronic hypertension requiring therapy, endocrine disease, and insulin-dependent diabetes. Participants completed two prenatal study visits. Visit 1 (V1) occurred at 16–29 weeks gestation and Visit 2 (V2) occurred at 22–39 weeks gestation, roughly corresponding to the 2nd and 3rd trimesters, respectively. This secondary analysis was conducted as part of the Environmental Influences on Child Health Outcomes (ECHO) PATHWAYS consortium.

2.2. Ethical approval

Institutional Review Board approval was obtained from the University of Tennessee Health Sciences Center (the primary site of data collection) and other participating institutions. All participants provided written informed consent prior to engaging in study activities.

2.3. Urine collection and phthalate metabolite measurement

Spot urine samples were collected in sterile, phthalate-free polypropylene containers at both prenatal visits. At the time of sample collection, specific gravity (SpG), a measure of urine dilution, was measured using a handheld refractometer. Samples were then frozen at -80°C until shipment (on dry ice) for phthalate metabolite analysis using previously described methods (Guo et al., 2014; Rocha et al., 2017). Enzymatic deconjugation, solid phase extraction, and high-performance liquid chromatography tandem mass spectrometry (HPLC-MS/MS) were used to assay a panel of 21 phthalate metabolites.

Blanks were included for several quality control samples including standard reference materials, procedural blanks, and matrix spikes. The analytical laboratory is part of the CDC Proficiency Testing Program (Biomonitoring Quality Assurance for State Program; BQASP) and German-External Quality Assurance Scheme (G-EQUAS) designed to optimize comparability and reliability of analyses across labs and between batches (Kannan et al., 2021).

The limits of detection (LODs) ranged from 0.012 to 0.304 ng/mL and only metabolites with $\geq 80\%$ of samples above the LOD were included in models for the analysis, resulting in 15 metabolites: mono-isobutyl phthalate (MiBP), monoethyl phthalate (MEP), mono-methyl phthalate (MMP), mono-n-butyl phthalate (MBP), mono-benzyl phthalate (MBzP), MCNP, mono-carboxy isooctyl phthalate (MCiOP), mono-carboxyisononyl phthalate (MCiNP), MCPP, MEHP, MEHHP, MEOHP, MECPP and mono(2-carboxymethylhexyl) phthalate (MCMHP), and phthalic acid (PA). Values below the LOD were imputed as $\text{LOD}/\sqrt{2}$ following convention (Hornung and Reed, 1990). Given the extensive evidence of di(2-ethylhexyl) phthalate (DEHP)'s endocrine-disrupting properties (reviewed in (Gore et al., 2015), following convention, we additionally calculated the molar sum of five di(2-ethylhexyl) phthalate (DEHP) metabolites (ΣDEHP : MEHP, MEHHP, MEOHP, MECPP, MCMHP) (Swan et al., 2015). Phthalate metabolite values were adjusted for urine dilution using the following formula: $[\text{phthalate metabolite}]_{\text{adj}} = [\text{phthalate metabolite}]_{\text{raw}} * [(\text{SpG}_{\text{median}} - 1) / (\text{SpG} - 1)]$ where $\text{SpG}_{\text{median}}$ refers to the median SG for the cohort (Boeniger et al., 1993). Phthalate metabolite concentrations were log-transformed in all analyses.

2.4. Blood collection and pCRH measurement

At both prenatal study visits maternal blood was collected using EDTA plasma separator tubes, and after processing, plasma was frozen at $-80\text{ }^{\circ}\text{C}$. Samples were shipped on dry ice to University of Newcastle, Australia. Using standard protocols, pCRH was measured (in pg/mL) by radioimmunoassay with an extraction recovery of 87% (Smith et al., 2009). The assay sensitivity was high compared to prior work with inter- and intra-assay coefficients of variation of 8.7% and 7.3%. pCRH values were log-transformed for analyses due to non-normality. In addition to considering V1 and V2 values independently, we calculated change in pCRH (ΔpCRH) as $(\log)V2\text{ pCRH} - (\log)V1\text{ pCRH}$.

2.5. Collection of covariate data

Pregnant women reported on demographic characteristics, health, and lifestyle behaviors during pregnancy. Variables of interest (selected a priori) included maternal age at enrollment, race and ethnicity (non-Hispanic White/non-Hispanic Black/Hispanic), pre-pregnancy body mass index (BMI; continuous), maternal education ($<$ high school/high school, GED, or technical school/college or greater), marital status (married or partnered/single), parity (parous/nulliparous), fetal sex, and gestational weeks at biospecimen collection. Gestational age was calculated based on last menstrual period and subsequently confirmed with ultrasound dating; in cases of discrepancies, the latter was used for final determination of gestational age. At each study visit, maternal active smoking was defined as cotinine levels at or above 200 ng/mL as measured in single spot urine samples. Based on prior analyses in this cohort, we additionally included childhood trauma (which was previously related to pCRH) in models, but did not include maternal psychosocial measures in pregnancy (which were not associated with pCRH) (Steine et al., 2020). Maternal history of childhood trauma was assessed using three items from the Traumatic Life Events Questionnaire (TLEQ): (1) physical abuse; (2) witnessing family violence while growing up; and (3) sexual abuse before age 13. A total childhood traumatic exposure types count was created based on these measures (0–3 range) (Kubany et al., 2000; Slopen et al., 2018). Finally, maternal gestational diabetes and gestational hypertension were determined by

participant report and confirmed by medical record abstraction. Both were included in the current analysis based on prior work in this cohort suggesting associations with pCRH (Steine et al., 2020).

2.6. Statistical analysis

CANDLE participants with phthalate metabolite and pCRH measurements at V1 and/or V2 were eligible for inclusion in the current analysis. Descriptive statistics (geometric mean, standard deviation, median, mix, max, $\%<$ LOD, percentiles, frequency) were conducted to examine the distribution of phthalate metabolite and pCRH concentrations (at each time point) as well as the covariates of interest. Bivariate analyses were used to examine the relationship between the variables of interest and histograms and scatterplots were created to visualize these associations.

We assessed three outcome measures: $\log(V1\text{ pCRH})$, $\log(V2\text{ pCRH})$, and ΔpCRH , equivalent to $\log(\text{pCRH})$ at V2 minus $\log(\text{pCRH})$ at V1. Our exposure was phthalate metabolite concentrations. Models predicting $\log(V1\text{ pCRH})$ included V1 phthalate metabolite concentrations, whereas models predicting $\log(V2\text{ pCRH})$ included V2 phthalate metabolite concentrations. For models predicting ΔpCRH between the two visits as the outcome, we included V1 values for gestational age, phthalate metabolites, and cotinine as well as the change in gestational age between the two study visits (weeks). In general, for time varying covariates (gestational weeks and cotinine) we included the values measured at the same visit as the outcome measures (e.g., V1 cotinine and V1 gestational age in models predicting V1 pCRH). In all multi-variable models, we adjusted for a set of covariates selected a priori based on the prior literature, particularly our recent work on determinants on pCRH in this cohort (Steine et al., 2020). These included: maternal age, race/ethnicity, marital status, education, parity, fetal sex, maternal urinary cotinine, maternal history of childhood trauma, gestational diabetes, gestational hypertension, and gestational age at sample collection.

Our primary models utilized weighted quantile sum (WQS) regressions to model the joint effect of all specific gravity-adjusted phthalate metabolites on the pCRH outcomes and their relative contributions to that mixture effect (Carrico et al., 2015). In WQS regression, each SG-adjusted phthalate metabolite is divided into quintiles, multiplied with a model-derived simplex vector of weights, and summed to generate a single weighted mixture index called the WQS which is then used as a predictor variable in a linear regression (Carrico et al., 2015). WQS regression is constrained to evaluate mixture effects in either a positive or negative direction, and both directions were evaluated for each outcome. In total, 1000 bootstraps were used to obtain stable mixture weight estimates for each WQS regression (Carrico et al., 2015). When <100 of the 1000 bootstrapped weights were associated with sum mixture coefficients in a given direction, we made note of this by lightening the plot colors for those estimates, and we recommend interpreting those results with caution. There may be no detectable mixture association in a given direction for some models, and in those cases the models will not return any estimates. The ‘‘HC0’’ Huber-White heteroskedasticity-consistent standard error sandwich estimator was used to calculate confidence intervals (White, 1980). WQS regressions are commonly performed using separate training and validation datasets to avoid high Type I error, though our own simulations show this also leads to a substantial loss of power [under review], which we avoided by using the full dataset for both training and validation. However, this can generate anti-conservative confidence intervals and p-values, and so we additionally applied a permutation test (Curtin et al., 2021) with 200 iterations to generate proper ‘‘confirmatory’’ p-values with a nominal Type I error rate that more accurately estimate WQS coefficient uncertainty (Day et al., 2021; Curtin et al., 2021), under review]. As permutation test p-values will always be higher than the original p-values, permutation tests were only applied to models with original p-values < 0.05 .

In addition to fitting mixtures models, secondarily, we employed the more conventional approach of considering the log-transformed, specific gravity-adjusted phthalate metabolites (and Σ DEHP) in individual linear regression models (one metabolite per model as well as a model with Σ DEHP as the main exposure measure). The same three outcomes (V1 pCRH, V2 pCRH, and Δ pCRH) were considered and we included the same set of covariates described above. We elected to fit individual linear regression models rather than hierarchical longitudinal models as we were concerned that the associations between phthalates and CRH might differ across the two visits, given the exponential rise in CRH that occurs across pregnancy (Moog et al., 2016; Smith et al., 2009). In the individual models, we additionally assessed the possibility of non-linearity by creating smooth plots of component plus residuals versus log-transformed, SG-adjusted phthalate metabolite concentrations by Visit.

Based on our observations regarding pCRH in prior work in this cohort (Steine et al., 2020), we additionally considered effect modification by fetal sex, gestational hypertension, gestational diabetes, and childhood trauma. To evaluate effect modification, we refit the WQS regression models while stratifying for each of the pregnancy complications individually, employing an approach in which random subsets of predictor variables are selected instead of bootstrapping observations for the weight estimation stage of the model (Curtin et al., 2021). This procedure, random subset WQS regression (WQS_{RS} regression), has an advantage over the bootstrap method when categorical covariates have few subjects in one or more strata. In this situation the random subset procedure is unaffected, but bootstrapping can result in some bootstrapped samples with no subjects in the rare stratum, causing an unidentified model. The random subset approach was selected here because there were small numbers of observations in some strata for the ethnicity covariate for observations having either pregnancy complication. As in the mixtures models based on the full cohort, we again did not split the data into training and validation datasets and then implemented the permutation test to maintain power while generating accurate p-values. For models considering interaction by fetal sex or childhood trauma, few significant interaction terms were observed, thus those analyses were not pursued further (Supplemental Table 1).

In our secondary models, we evaluated effect modification in the models with individual phthalate metabolites by including interaction terms (e.g., phthalate*gestational diabetes). We reparameterized the individual phthalate metabolite models to obtain effect estimates and 95% confidence intervals (CIs) for all women with and without each of these pregnancy complications. All analyses were conducted using R statistical software. WQS models were constructed using the gWQS package (version 3.0.4) (Renzetti et al., 2021).

3. Results

3.1. Descriptive statistics

In total, 1483 women were enrolled in the study at V2 and had a live birth. pCRH concentrations were measured in samples from 1303 women and phthalate metabolite concentrations were measured in 1173, resulting in a total of 1018 women with complete data to contribute to the current analyses at V1 and 1014 at V2. On average, women were 26.4 ± 5.5 years old with a pre-pregnancy BMI of 27.9 ± 7.7 kg/m² (Table 1). Most participants identified as non-Hispanic Black (61.4%), with the remainder identifying as Non-Hispanic White (32.0%) or other races/ethnicities (6.6%). The majority (56.2%) had a high school, GED, or technical school education. <10% of women were smokers (6.4% and 9.9% in V1 and V2, respectively) and 39.9% of women were nulliparous. During study participation, 59 (5.8%) and 102 (10.0%) women developed gestational diabetes and gestational hypertension, respectively.

The phthalate metabolites of interest were detectable in the vast majority of women, with 12 of 15 metabolites detectable in >95% of participants (not shown). At both visits, median MEP concentrations

Table 1

Characteristics of CANDLE mother-child dyads (n = 1018).

| Characteristics (continuous) | Mean \pm SD | N (%) |
|--|----------------|------------|
| Maternal age (years) | 26.4 \pm 5.5 | |
| Pre-pregnancy BMI (kg/m ²) | 27.9 \pm 7.7 | |
| Gestational age at Visit 1 (V1; weeks) | 23.0 \pm 3.0 | |
| Gestational age at Visit 2 (V2; weeks) | 31.8 \pm 1.7 | |
| Change in gestational age (V2-V1; weeks) | 8.9 \pm 7.3 | |
| Maternal Childhood Traumatic Life Events | 0.53 \pm 0.8 | |
| Physical punishment | | 82 (8.1) |
| Exposure to family violence | | 277 (27.2) |
| Sexual abuse | | 184 (18.1) |
| Characteristics (categorical) | | |
| Maternal race/ethnicity | | |
| Non-Hispanic White | | 326 (32.0) |
| Non-Hispanic Black | | 625 (61.4) |
| Hispanic | | 67 (6.6) |
| Highest level of maternal education | | |
| <High School | | 107 (10.5) |
| High School/GED/Technical School | | 572 (56.2) |
| College or Higher | | 339 (33.3) |
| Nulliparous | | 436 (39.9) |
| Cotinine detected in V1 urine | | 65 (6.4) |
| Cotinine detected in V2 urine | | 101 (9.9) |
| Gestational diabetes | | 59 (5.8) |
| Gestational hypertension | | 102 (10.0) |
| Fetal sex- female | | 510 (49.9) |

were highest out of all metabolites measured (V1: 114 ng/mL, V2: 103 ng/mL) and median MHPP concentrations were the lowest (V1: 1.02 ng/mL, V2: 0.35 ng/mL) (Table 2). Comparing women with and without gestational diabetes, levels of phthalate metabolites were similar at Visit 1, but tended to be lower (for 13 of 15 metabolites) among women with gestational diabetes at Visit 2. By contrast, women with gestational hypertension tended to have higher concentrations of phthalate metabolites than women without gestational hypertension, particularly at Visit 2 (Table 2). pCRH levels were considerably higher at V2 compared to V1 (median V1 pCRH: 37.6 pg/mL, V2 pCRH: 235.2 pg/mL) (Table 2). pCRH concentrations were higher among women with gestational diabetes and gestational hypertension compared to women without those complications, with the differences increasing dramatically by Visit 2.

3.2. Multivariable models: Full cohort

In covariate-adjusted WQS mixtures models using data from the full cohort, we observed a positive association between the phthalate mixture and pCRH at Visit 1 ($\beta = 0.07$; 95% CI: 0.02, 0.11; permutation test p (PTp) = 0.14; Fig. 1). By contrast, at Visit 2, there was an inverse association between the phthalate mixture and pCRH ($\beta = -0.08$; 95% CI: -0.14, -0.02; PTp = 0.06) with high weights for MEP and MCINP. Neither association was statistically significant after the permutation test. In individual metabolite models, at V1, MBP was associated with significantly higher pCRH ($\beta = 0.07$; 95% CI: 0.004, 0.13). While no other significant associations were observed at that timepoint, consistent with the results of the mixtures models, estimates were mostly positive (Supplemental Table 2). At V2, we observed a positive association between MPP and pCRH concentrations ($\beta = 0.04$; 95% CI: 0.01, 0.08) and a trend towards an inverse association between MEP and pCRH ($\beta = -0.03$; 95% CI: -0.07, 0.004); no associations with other metabolites were noted, however most estimates were in the negative direction. At both visits the estimate for the phthalate mixture was as strong or stronger than the results of any individual metabolite alone. Analyses examining the change in pCRH from V1 to V2 showed no associations with individual phthalate metabolite concentrations. Assessment of non-linearity (to examine potentially non-monotonic effects of phthalates on pCRH) suggested predominantly linear patterns with some evidence of change in slope at the highest concentrations ($\geq 95^{\text{th}}$ percentile) for the DEHP metabolites at V1 and for MCINP and MIBP at V2 (not shown).

Table 2

Median specific gravity adjusted phthalate metabolite¹ and pCRH concentrations by study visit, gestational hypertension (GHTN; n = 102), and gestational diabetes (GDM; n = 59).

| | VISIT 1 (16–29 weeks; n = 1018) | | | | | VISIT 2 (22–39 weeks; n = 1014) | | | | |
|--------------------------------------|---------------------------------|--------------|--------------|--------------|--------------|---------------------------------|---------------|---------------|---------------|---------------|
| | Total cohort | No GHTN | GHTN | No GDM | GDM | Total cohort | No GHTN | GHTN | No GDM | GDM |
| Phthalate metabolites (ng/mL) | | | | | | | | | | |
| MBP | 29.80 | 29.66 | 32.13 | 29.87 | 26.22 | 16.06 | 15.67 | 18.29 | 16.04 | 13.55 |
| MBzP | 19.35 | 18.68 | 21.20 | 18.98 | 18.67 | 10.40 | 10.20 | 12.79 | 10.48 | 9.17 |
| MCINP | 3.06 | 3.12 | 2.85 | 3.06 | 3.89 | 0.48 | 0.47 | 0.59 | 0.48 | 0.38 |
| MCIOP | 11.84 | 12.21 | 11.44 | 11.84 | 12.81 | 2.21 | 2.16 | 2.59 | 2.22 | 2.01 |
| MCP | 1.95 | 1.96 | 1.94 | 1.96 | 1.78 | 1.43 | 1.40 | 1.77 | 1.43 | 1.28 |
| MCMHP | 16.32 | 16.32 | 16.65 | 16.31 | 16.68 | 6.24 | 6.13 | 6.47 | 6.26 | 4.77 |
| MECPP | 17.21 | 16.89 | 19.69 | 17.28 | 17.96 | 11.90 | 11.65 | 13.37 | 11.90 | 9.19 |
| MEHHP | 26.18 | 26.12 | 27.33 | 26.22 | 26.40 | 8.27 | 8.12 | 9.60 | 8.38 | 5.99 |
| MEHP | 7.01 | 6.97 | 7.92 | 7.09 | 5.27 | 2.31 | 2.30 | 2.11 | 2.33 | 1.65 |
| MEOHP | 12.50 | 12.46 | 13.59 | 12.56 | 12.60 | 6.28 | 6.19 | 6.94 | 6.33 | 4.43 |
| MEP | 122.08 | 119.00 | 155.15 | 122.60 | 99.52 | 111.84 | 110.72 | 116.17 | 113.76 | 61.14 |
| MHPP | 1.10 | 1.12 | 1.01 | 1.10 | 1.12 | 0.37 | 0.36 | 0.41 | 0.37 | 0.36 |
| MiBP | 11.96 | 11.83 | 11.70 | 11.96 | 10.57 | 7.74 | 7.59 | 8.73 | 7.76 | 5.89 |
| MMP | 4.95 | 5.05 | 4.92 | 5.14 | 2.38 | 2.39 | 2.36 | 2.82 | 2.40 | 2.11 |
| PA | 59.53 | 58.53 | 72.75 | 59.46 | 65.27 | 70.12 | 68.06 | 81.06 | 70.41 | 58.86 |
| pCRH (pg/mL) | 37.60 | 37.97 | 40.28 | 37.81 | 41.81 | 235.20 | 229.90 | 334.88 | 230.96 | 313.85 |

¹ LODs for phthalate metabolites range from 0.012 to 0.137.

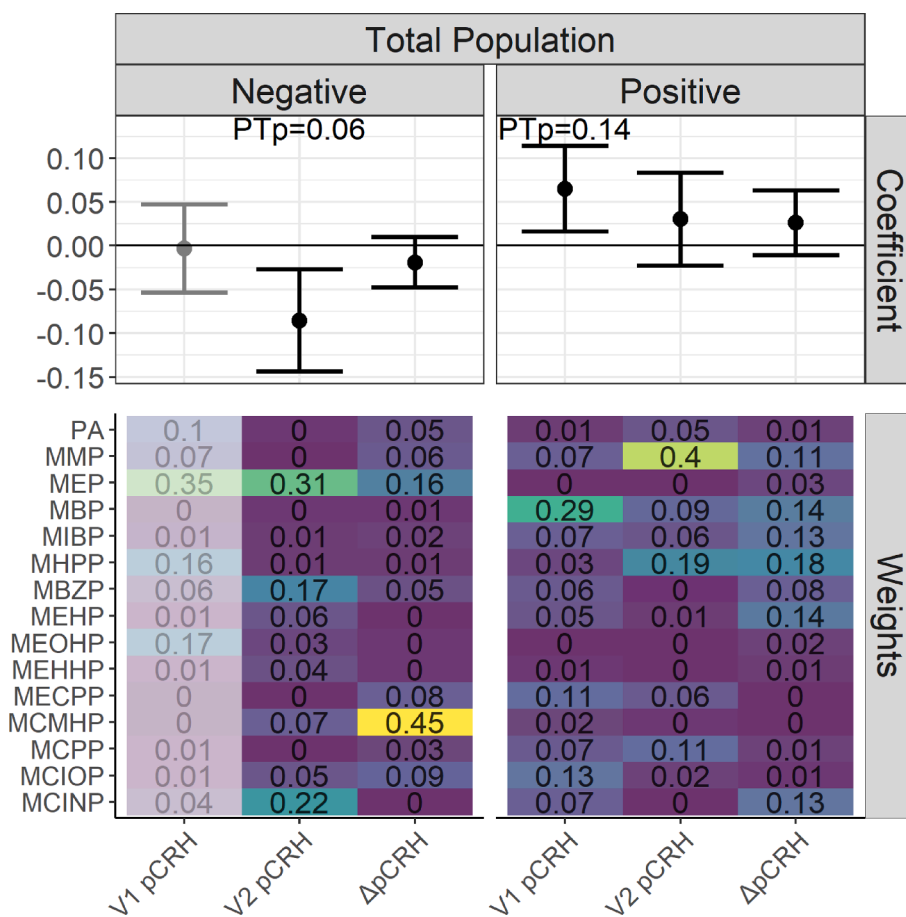


Fig. 1. WQS regression coefficients and weights for associations between phthalate mixtures and log(pCRH) in pg/mL^{1,2}. The forest plots show WQS regression means and 95% CIs in the negative and positive directions. Permutation test p-value (PTp) indicates p-values after the application of the permutation test. WQS weights for all phthalate metabolites are indicated on the heat maps³. (1 Models adjusted for gestational age at sample collection, cotinine, maternal age, maternal race and ethnicity, marital status, fetal sex, maternal education, pre-pregnancy BMI, parity, gestational diabetes, gestational hypertension, and maternal childhood traumatic life events. Models examining the outcome ΔpCRH are additionally adjusted for the change in gestational age between the visits. 2 Exposure is Visit 1 phthalate metabolite concentrations for the outcome Visit 1 log(pCRH). Exposure is Visit 2 phthalate metabolite concentrations for the outcomes Visit 2 log(pCRH) and ΔpCRH. 3 The WQS regression coefficient and weight estimates in the negative direction for the V1 pCRH outcome are de-emphasized since these model results were derived from < 100 (<10%) of the total bootstrap iterations within the WQS regression. These bootstrap iterations in a given direction are used to derive mixture weights and to compile to WQS mixture index, and so estimates based on only a few iterations in the desired direction may be unstable and should be interpreted with caution).

3.3. Stratified analyses: Gestational diabetes

Stratified analyses indicated differing patterns of association in relation to gestational diabetes status. In WQS regression models, among women who developed gestational diabetes in pregnancy, a significant inverse mixture association with pCRH was observed at V2 ($\beta = -0.35$; 95% CI: $-0.50, -0.19$; $PTp < 0.005$) with similarly high weights for MBP, MEOHP, MEHHP, and MCIOP (Fig. 2). By contrast, among women

without gestational diabetes, we observed a positive mixture association at V1 ($\beta = 0.07$; 95%CI: 0.02, 0.11; $PTp = 0.05$) with high weights for MBP, MCIOP, and MCP, although this was nonsignificant after the permutation test (Fig. 2). When we considered the change in pCRH between the two visits, we observed a strong positive association among women with gestational diabetes ($\beta = 0.16$; 95% CI: 0.07, 0.25; $PTp = 0.02$) that was driven by the DEHP metabolites MEHHP, MECPP, and MEHP. By contrast, among women without gestational diabetes we

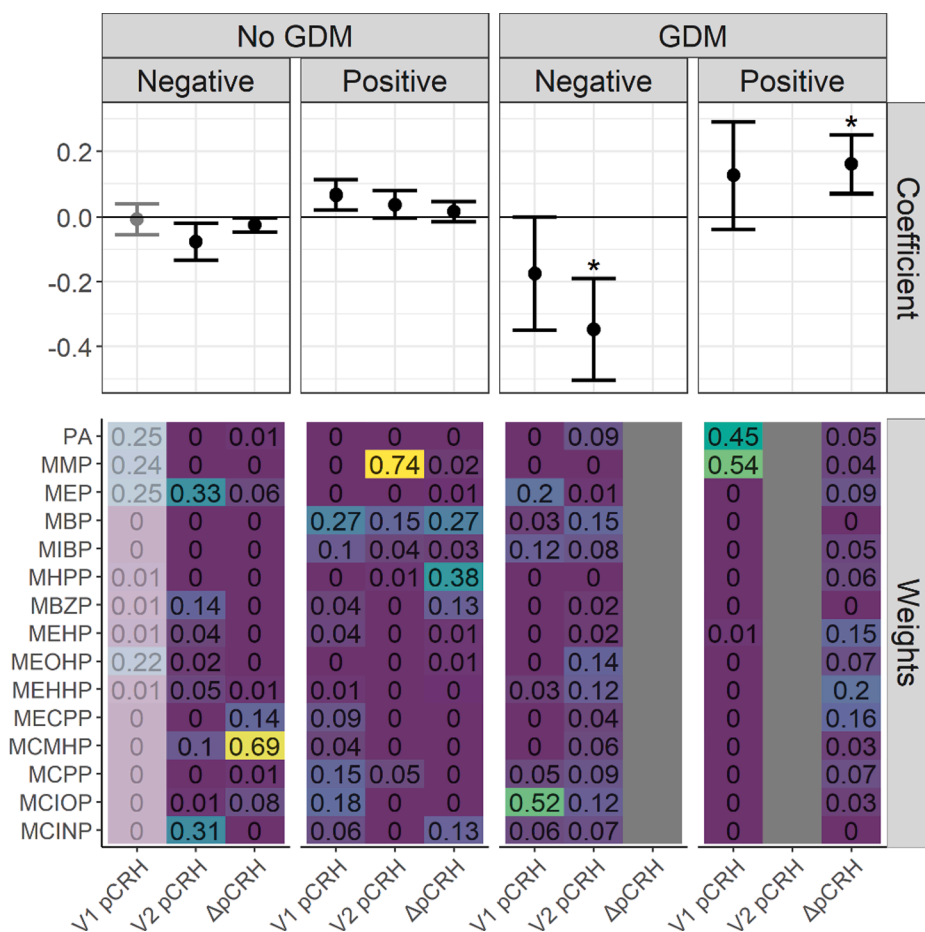


Fig. 2. WQS regression coefficients and weights for associations between phthalate mixtures and log(pCRH) in models stratified by gestational diabetes (GDM) status (in pg/mL)^{1,2}. The forest plots show WQS regression means and 95% CIs in the negative and positive directions. Asterisks denote permutation test p-values < 0.05. WQS weights for all phthalate metabolites are indicated on the heat maps³. (1 Models adjusted for gestational age at sample collection, cotinine, maternal age, maternal race and ethnicity, marital status, fetal sex, maternal education, pre-pregnancy BMI, parity, gestational diabetes, gestational hypertension, and maternal childhood traumatic life events. Models examining the outcome ΔpCRH are additionally adjusted for the change in gestational age between the visits. 2 Exposure is Visit 1 phthalate metabolite concentrations for the outcome Visit 1 log(pCRH). Exposure is Visit 2 phthalate metabolite concentrations for the outcomes Visit 2 log(pCRH) and ΔpCRH. 3 For participants with diabetes, the WQS regression for ΔpCRH in the negative direction and for V2 pCRH in the positive direction found no bootstrapped mixture coefficients in the desired direction and therefore returned no estimate for those respective directions and outcomes, and therefore no estimates are presented for those models on the plots. Some WQS regression estimates (e.g., the negative mixture association with V1 pCRH for participants without diabetes) are de-emphasized since these model results were derived from < 100 (<10%) of the total bootstrap iterations within the WQS regression. These bootstrap iterations in a given direction are used to derive mixture weights and to compile to WQS mixture index, and so estimates based on only a few iterations in the desired direction may be unstable and should be interpreted with caution).

observed a weak inverse association ($\beta = -0.02$; 95 %CI: $-0.05, -0.002$; PTp = 0.46) with MCMHP most heavily weighted (Fig. 2).

Results of secondary models examining interactions between individual phthalate metabolite concentrations and gestational diabetes were similar (but not identical) to WQS results (Supplemental Fig. 1). In individual metabolite models, we observed no evidence of interactions at V1 (not shown). At V2, interactions with gestational diabetes were observed for a number of phthalate metabolites including MBP, MEHHP, MEOHP, MIBP, and phthalate acid as well as ΣDEHP. Among women with gestational diabetes, all associations between phthalate metabolites and pCRH at V2 were inverse and tended to be stronger than those observed in women without gestational diabetes. Among women with gestational diabetes, significant inverse associations were also noted for other metabolites at V2 including MEHHP, MEOHP, MIBP, and ΣDEHP. Among women without gestational diabetes, the only associations observed were a negative association between MBzP and pCRH ($\beta = -0.04$, 95% CI: $-0.08, -0.01$) and a positive relationship between MMP and pCRH ($\beta = 0.05$, 95% CI: $0.01, 0.09$). When we examined ΔpCRH from V1 to V2, results were similar, with phthalate*gestational diabetes interactions noted for a number of metabolites, including MCPP, MECPP, MEHHP, MEHP, MEOHP, MCMHP, and MMP, as well as ΣDEHP. With the exception of MBP, among women with gestational diabetes, all metabolites were positively associated with ΔpCRH indicating higher phthalate metabolite concentrations were associated with a greater rise in pCRH between the two visits. These associations were significant for ΣDEHP ($\beta = 0.139$, 95% CI: $0.003, 0.276$) as well as the DEHP metabolites, MECHP, MEHP, and MEOHP. Among women without gestational diabetes, the direction of association between phthalate metabolites and ΔpCRH was variable and associations were weaker and non-significant for all metabolites.

3.4. Stratified analyses: Gestational hypertension

Stratified analyses also suggested different patterns of association in relation to gestational hypertension. In WQS regression models, among women who developed gestational hypertension in pregnancy, we observed positive associations with pCRH at both V1 ($\beta = 0.20$; 95% CI: $0.03, 0.36$; PTp = 0.16) with high weights for MCPP, MCIOP and MBzP; as well as V2 ($\beta = 0.42$; 95% CI: $0.19, 0.64$; PTp = 0.005) with high weights for MMP, MCIOP, MIBP, and MECPP (Fig. 3). However, we observed an inverse association with ΔpCRH ($\beta = -0.17$; 95% CI: $-0.29, -0.06$; PTp = 0.06) with high weights for MBP, MCMHP, and MEP. Only the positive association at V2 remained statistically significant after the permutation test. By contrast, among women without gestational hypertension, associations were weakly positive at V1 ($\beta = 0.06$; 95% CI: $0.01, 0.11$; PTp = 0.09) and negative at V2 ($\beta = -0.09$; 95% CI: $-0.14, -0.04$; PTp = 0.01) (Fig. 3). Only the estimate for the negative direction at V2 was statistically significant after the permutation test. MBP was the strongest contributor to the positive associations observed at V1 whereas MCINP, MEP, and MEHHP were most heavily weighted at V2.

In linear regression models examining interactions between individual phthalate metabolites and gestational hypertension, we again observed no associations at V1 (not shown), with associations apparent at V2 (Supplemental Fig. 2). Interactions between gestational hypertension and phthalates were observed for most metabolites at V2, particularly MBzP, MCINP, MCIOP, ΣDEHP, MECPP, MEHHP, MEOHP, MIBP, and MMP. Among women with gestational hypertension, all phthalate metabolites were positively associated with pCRH levels at V2, whereas most associations were inverse and non-significant among women without gestational hypertension. Among women with gestational hypertension, significant positive associations were also noted for

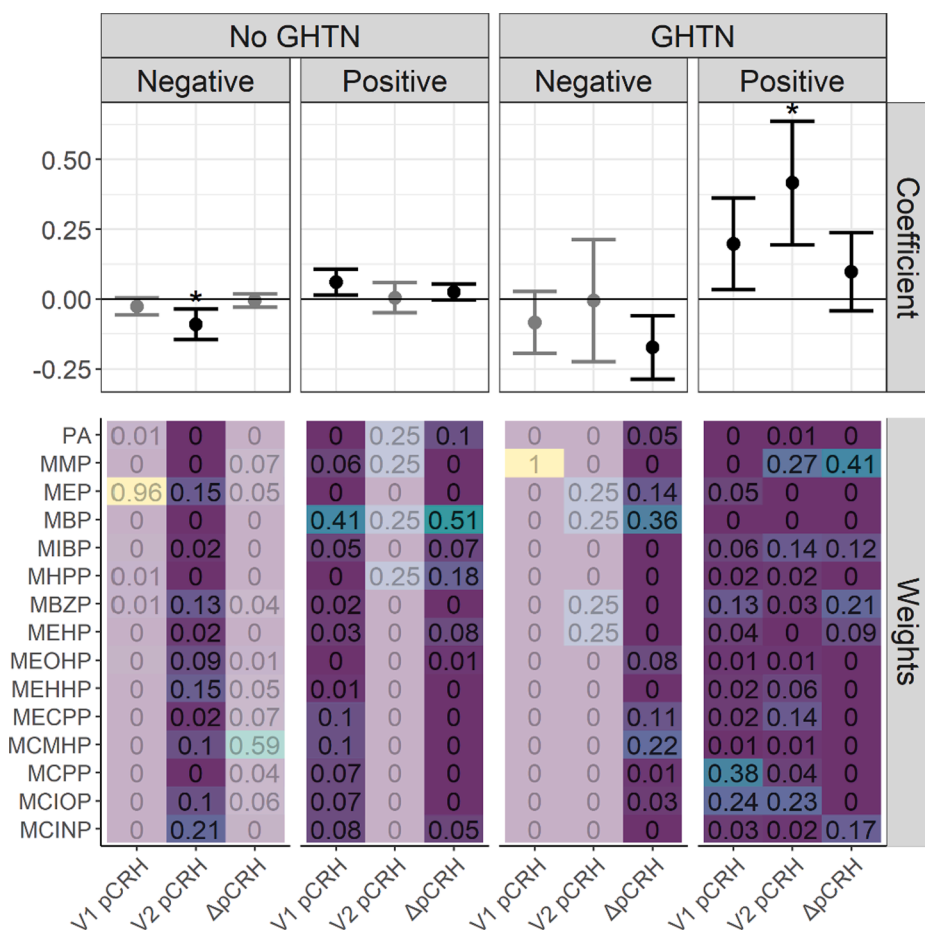


Fig. 3. WQS regression coefficients and weights for associations between phthalate mixtures and log(pCRH) in models stratified by gestational hypertension (GHTN) status (in pg/mL)^{1,2}. The forest plots show WQS regression means and 95% CIs in the negative and positive directions. Asterisks denote permutation test p-values < 0.05. WQS weights for all phthalate metabolites are indicated on the heat maps³. (1 Models adjusted for gestational age at sample collection, cotinine, maternal age, maternal race and ethnicity, marital status, fetal sex, maternal education, pre-pregnancy BMI, parity, gestational diabetes, gestational hypertension, and maternal childhood traumatic life events. Models examining the outcome ΔpCRH are additionally adjusted for the change in gestational age between the visits. 2 Exposure is Visit 1 phthalate metabolite concentrations for the outcome Visit 1 log(pCRH). Exposure is Visit 2 phthalate metabolite concentrations for the outcomes Visit 2 log(pCRH) and ΔpCRH. 3 Some WQS regression estimates (e.g., the negative mixture association with V1 pCRH for participants without GHTN) are de-emphasized since these model results were derived from <100 (<10%) of the total bootstrap iterations within the WQS regression. These bootstrap iterations in a given direction are used to derive mixture weights and to compile to WQS mixture index, and so estimates based on only a few iterations in the desired direction may be unstable and should be interpreted with caution.)

a number of other metabolites at V2, including MCIOP, MECPP, MEHHP, MIBBP, and MMP, and as well as ΣDEHP. Among women without gestational hypertension, V2 pCRH was significantly and inversely associated with several metabolites including MBzP, MEHHP, and MEP. Results from models examining the interaction between ΔpCRH and gestational hypertension showed few significant interactions, with the exception phthalic acid. Among women with gestational hypertension, most (but not all) phthalate metabolites showed negative associations with ΔpCRH, suggesting a lower rise between the two visits, though results were only significant for phthalic acid. Among women without gestational hypertension, the direction of association differed by metabolite and no significant associations were observed.

4. Discussion

In this analysis of 1018 participants from the CANDLE pregnancy cohort, we evaluated associations between gestational phthalate exposures and pCRH concentrations at two timepoints, mid and late pregnancy. Overall, we observed that phthalate mixtures were associated with higher pCRH in mid-pregnancy, but lower pCRH later in pregnancy. Although few associations were observed when we considered the individual phthalate metabolites in separate models, MBP appeared to drive associations at the first visit, while MEP drove associations in the second visit. When we subsequently considered pregnancy complications in our models, effect modification was evident. Among women with gestational diabetes, phthalate mixtures were associated with lower pCRH concentrations at each timepoint (particularly in late pregnancy), but a steeper rise in pCRH. Among women with gestational hypertension, phthalate mixtures were associated with higher pCRH at

both timepoints (again, particularly in late pregnancy), but also a lower rise in pCRH between the visits. Analyses considering individual metabolites further corroborated these results. Among women who did not have pregnancy complications, phthalates were associated with lower pCRH in late pregnancy (V2), though the associations were weaker than those observed among women with complications. In general, associations between phthalates and pCRH among women with pregnancy complications tended to be more pronounced in late pregnancy (V2) than in mid-pregnancy (V1), which may reflect increasing production of pCRH and/or greater severity of pregnancy complications in late pregnancy.

Many epidemiological studies have examined associations between phthalates and preterm birth with most, but not all, reporting that DBP, diisobutyl phthalate (DiBP), butyl benzyl phthalate (BBzP), and DEHP may be associated with adverse outcomes (Ferguson et al., 2019; Cantonwine et al., 2016; Meeker et al., 2009; Zhong et al., 2021; Ruiz et al., 2016; Ferguson et al., 2014; Ferguson et al., 2014; Ferguson et al., 2019; Gao et al., 2019). Our results suggest that phthalate exposure may impact pCRH production and that those impacts may vary across pregnancy, but the mixed directionality of these associations does not clearly support the hypothesis that phthalates contribute to preterm birth through the pCRH pathway. Other pathways of interest include oxidative stress, inflammation, and other hormones; research on these mechanisms is underway (reviewed in Ferguson and Chin, 2017). To our knowledge, no study of phthalates and preterm birth has specifically considered women with pregnancy complications, whose risks and vulnerability to phthalates may differ from the general population.

Previous epidemiological work on phthalates and pCRH is extremely limited. In PROTECT, a study of 676 pregnant women in Puerto Rico, phthalate metabolites and pCRH were measured at two visits at 16–20

and 24–28 weeks gestation (Cathey et al., 2019). In linear mixed models examining phthalate metabolites individually, multiple metabolites (including MCNP, MCCP, MECPP, MEHHP, and MEOHP) were associated with lower pCRH concentrations, with the strongest associations observed for the DEHP metabolites. For example, an interquartile (IQR) increase in MECPP was associated with 18% lower pCRH levels; associations were significant, but slightly weaker, for other DEHP metabolites such as MEHHP and MEOHP. In general, associations were stronger later in gestation. In our study, results from the second visit (at 22–39 weeks) were generally consistent with the PROTECT findings, although MEP appeared to drive our association rather than the DEHP metabolites. However, our results earlier in pregnancy suggesting positive associations diverged from their findings.

Of note, pCRH concentrations differed considerably across the two studies, with median pCRH concentrations of 82.4 and 86.6 pg/mL at the two PROTECT visits as compared to 37.6 and 235.2 pg/mL at the two CANDLE visits. This difference may partially reflect differences in timing of the visits, however the lack of a rise in pCRH between the two visits in PROTECT is surprising given the documented exponential rise in pCRH during mid-late pregnancy (Frim et al., 1988; Vale et al., 1981; Campbell et al., 1987). Notably, PROTECT pCRH values were based on serum samples assayed using enzyme linked immunoassay (ELISA), whereas CANDLE analyses were performed on plasma samples assayed using radioimmunoassay (RIA). Plasma has been proposed as the preferred matrix and RIA, which is more widely used in the pCRH literature, may provide greater sensitivity than ELISA (Latendresse and Ruiz, 2008). To date, evidence directly comparing the two analytic approaches is limited. However consistent with the differences observed between our results and those of PROTECT, in a recent comparison of assay types, we observed a clear increase in pCRH across gestation when RIA was used, while concentrations were flat over time when ELISA was used (Herrera et al., 2021). Overall, the numerous methodological differences may contribute to differences in results in these two studies. Unfortunately, no information was provided on pregnancy complications (including gestational diabetes and gestational hypertension) among PROTECT participants, precluding that comparison.

To our knowledge, no other epidemiological study has examined this association, though there is at least one complementary *in vitro* study on this topic (Wang et al., 2016). Wang et al. administered MEHP at doses from 1 to 150 μM to purified primary cytotrophoblasts from healthy term human placentas and measured pCRH expression. No changes were observed at doses below 100 μM , however at 100 and 150 μM , pCRH was upregulated in a dose-dependent manner. Knockdown experiments additionally identified NF- κB inducing kinase (NIK), a signaling component of the NF- κB pathway, as central to MEHP's upregulation of pCRH. These results contrast to those reported in the PROTECT study and to some extent, the current results, both of which suggested lower pCRH in relation to DEHP metabolite exposure in healthy pregnant women. However, the doses used by Wang et al were higher than those observed in human studies (including CANDLE) and reflected acute exposures (24 h) in term placental tissue, making them less directly applicable to human studies based on chronic exposures starting well before parturition. At present, we know of no *in vitro* or animal studies examining chronic, low-dose exposure to phthalates (similar to that experienced by humans) in relation to pCRH.

That pCRH may be vulnerable to endocrine disruption is further supported by several recent epidemiological studies looking at other environmental chemical exposures. Additional work in the PROTECT cohort examined 12 phenols and parabens in relation to pCRH concentrations in pregnancy (Aker et al., 2019). An IQR increase in Bisphenol S (a Bisphenol A substitute increasingly used in plastics manufacture) was associated with an 11.35% decrease in pCRH (95% CI: -18.71 , -3.33) with stronger associations in early versus late pregnancy. By contrast, a IQR increase in triclosan, an antibacterial and antifungal chemical widely used in cleansers, was associated with a 9.20% increase in pCRH (95% CI: -0.97 , 20.42). Similarly, in a

California pregnancy cohort, second trimester pCRH was examined in relation to exposure to per- and poly-fluoroalkyl substances, a class of persistent synthetic chemicals found in consumer products such as cookware, food, drinking water, and clothing (Eick et al., 2021; Sunderland et al., 2019). In that study, an IQR increase in perfluorononanoic acid was associated with higher pCRH ($\beta = 5.17$, 95% CI: 1.79, 8.55) and weaker associations were also observed for perfluorooctanoic acid ($\beta = 3.62$, 95% CI: -0.42 , 7.66). Interestingly, in stratified analyses, associations were stronger among women who also reported higher levels of non-chemical stressors (e.g., depression, food insecurity, and financial strain) compared to women who did not report such stressors. By contrast, in our study, we did not observe an interaction between phthalate exposures and stressors (in the form of childhood trauma history) on CRH. Notably, the recall of childhood trauma was limited to items on threatening experiences (e.g. sexual and physical violence), but we did not assess deprivation adversity (as the California study did). It is possible that phthalate*stressor interactions would have been observed with the use of a more extensive childhood trauma scale. Another notable difference was that the California study examined current stressors (but not childhood trauma) whereas we did not include current stressors in models given their lack of association with CRH in our prior work (Steine et al., 2020).

Among CANDLE participants, concentrations of many phthalate metabolites were somewhat higher than those reported in other pregnancy cohorts recruited contemporaneously. For instance, median levels of MBzP were 18.81 and 9.65 ng/mL (at Visits 1 and 2, respectively) in CANDLE, compared to 3.10 ng/mL in the multi-center U.S. pregnancy cohort TIDES (recruitment from 2010 to 2012) and 7.0 ng/mL in the Boston-based LIFECODES study (among participants recruited 2006–2008) (Swan et al., 2015; Bellavia et al., 2017). Such differences in exposure may be geographic and may also reflect higher phthalate exposures among non-White women, as have been reported elsewhere (James-Todd et al., 2016; Bloom et al., 2019). Consistent with those previously reported disparities, in CANDLE, concentrations of some phthalate metabolites (most notably MBP and MEP, found in personal care products) were higher among Black participants compared to White participants (not shown). Some evidence suggests that products marketed towards women of color may contain high concentrations of toxic chemicals including endocrine disruptors, contributing to disproportionate burden of chemical exposures (Zota and Shamasunder, 2017). Racial/ethnic, geographic, and socioeconomic differences in diet, particularly consumption of processed and fast foods, may also contribute to disparities in phthalate exposures (Buckley et al., 2019; Zota et al., 2016; Martínez Steele et al., 2020).

A novel finding emerging from our analyses was the notable difference in associations between women with and without pregnancy complications in both mixtures and individual metabolite models. A growing number of studies have examined prenatal phthalate exposure as a potential contributor to hypertensive disorders of pregnancy and gestational diabetes. While evidence implicating phthalates in the etiology of gestational diabetes is fairly consistent and compelling (Bellavia et al., 2017; James-Todd et al., 2016; Shaffer et al., 2019; Zukin et al., 2021; Fisher et al., 2018); results of studies on associations with gestational hypertensive disorders have been more mixed (Warembourg et al., 2019; Werner et al., 2015; Cantonwine et al., 2016; Phillips et al., 2019). Largely ignored in the literature thus far, however, the extent to which women with pregnancies complicated by these disorders may be more vulnerable to further physiologic dysregulation by environmental chemical exposures. Theoretically, pregnancy complications represent physiologic stressors, and the additional burden of chemical exposures may result in greater dysregulation (for instance, of pCRH production). In this study, we were unable to examine the timing of urine collection relative to the diagnosis of pregnancy complications. Given that GDM is typically diagnosed through glucose challenge tests administered between 24 and 28 weeks gestation, it is probable that the second urine collection (at mean GA 31.8 ± 1.7 weeks) occurred after GDM diagnosis

(if any), and notably, among women with GDM, associations were stronger at the second visit relative to the first. Diagnosis of GHTN, which is more temporally variable, could have occurred before or after Visit 2. As was the case with GDM, among women with GHTN, associations between phthalate metabolites and pCRH were again stronger at the second visit, this time in the positive direction. An alternative explanation for these results is that dysregulation of pCRH contributes to pregnancy complications (as some studies suggest) and that environmental exposures may further exacerbate those risks.

In our study, among women with gestational hypertension, phthalate mixtures were associated with higher pCRH levels across pregnancy but a lower pCRH rise between the two visits. By contrast, in women without gestational hypertension, associations were weaker and in late pregnancy, the phthalate mixture was associated with lower pCRH. Women with gestational hypertension also tended to have higher median concentrations of most phthalates and higher pCRH at both timepoints. Indeed, multiple studies have indicated that women with hypertensive disorders of pregnancy may have higher pCRH levels, but lower CRH binding protein levels than compared to women with uncomplicated pregnancies (Laatikainen et al., 1991; Perkins et al., 1993; Perkins et al., 1995; Karteris et al., 2003; Purwosunu et al., 2007; Liapi et al., 1996). High pCRH levels have also been associated with increased placental resistance as assessed via uterine artery pulsatility and arterial resistance (Harville et al., 2008). More recent work has found increased methylation of the gene encoding CRH binding protein in women with early (but not late) onset pre-eclampsia, compared to women with uncomplicated pregnancies (Hogg et al., 2013). Elevated pCRH may contribute to preeclampsia risk, furthermore, through dysregulation of the placental nitric oxide/cGMP pathway (Clifton et al., 1995; Karteris et al., 2005; Makriganakis et al., 2018).

We also observed associations in women with gestational diabetes. Phthalate mixtures were associated with lower pCRH levels at both visits (with stronger associations observed at the second visit, which would be after the time of a typical gestational diabetes diagnosis), as well as a larger rise in pCRH between visits. In this sample, women with gestational diabetes tended to have lower median phthalate metabolite concentrations compared to women without gestational diabetes, particularly in late pregnancy. In contrast to the literature on hypertensive disorders and pCRH, little epidemiological work has examined relationships between pCRH and gestational diabetes, though in this CANDLE sample, pCRH levels were higher among women with gestational diabetes compared to those without gestational diabetes. Interestingly, *in vitro* work in trophoblast cells showed that pCRH modulates the expression of glucose transporters, GLUT1 and GLUT3, suggesting a role of pCRH in glucose activity during pregnancy (Gao et al., 2012). Work in mouse models additionally suggests that blocking CRH receptor signaling in pregnant dams results in impaired glucose tolerance, without changes in insulin sensitivity or β -cell proliferation (Simpson et al., 2020). It has also been proposed that by stimulating cortisol production, pCRH may mobilize glucose in the maternal bloodstream, promoting fetal growth (Gangestad et al., 2012). This is supported by epidemiological work indicating positive associations between pCRH and cord blood glucose (Valsamakis et al., 2020). With limited understanding of the associations between pCRH and pregnancy complications, it is difficult to evaluate how phthalates may fit in. Given the very limited work in this area, there is a need for additional work specifically designed to evaluate the impact of environmental exposures in women with complicated pregnancies.

Our study has several notable strengths. The CANDLE cohort is large and this analysis of 1018 women included 625 Black women, a group that is typically under-represented in pregnancy cohort studies, yet who may experience higher phthalate exposures due to lifestyle and consumer goods (James-Todd et al., 2016; Chan et al., 2021). The cohort is also socioeconomically diverse and resides in the Southern U.S., a geographic area that has been underrepresented in environmental epidemiology cohorts. We assessed mixtures of phthalates at two time

points through WQS regression, allowing us to simultaneously quantify numerous correlated metabolites. This increasingly popular mixtures approach may better approximate real-life exposure than the traditional approach of considering each metabolite individually and in general, the associations observed were stronger in the mixtures models compared to the individual phthalate models. In addition, we present novel results suggesting effect modification in relation to common pregnancy complications (gestational diabetes and gestational hypertension). While many studies have examined phthalates as potential contributors to these disorders (e.g. James-Todd et al., 2016; Shaffer et al., 2019; Fisher et al., 2018; Philips et al., 2019), we know little about whether these complications may also enhance vulnerability to endocrine disruption, as our results may suggest. Finally, very little is known about pCRH as a target of endocrine disruption and this research adds to that small literature. We used highly precise and reliable gold standard assays to measure pCRH in mid- and late-pregnancy and our results suggest a need for further research on this important, understudied hormone.

At the same time, we note several limitations. Phthalates are non-persistent chemicals and their half-lives in the human body are several hours (Koch et al., 2006). Thus, a single spot urine collection represents only recent exposures and may not accurately capture exposure levels across longer time spans, such as a trimester of pregnancy, resulting in the potential for exposure misclassification and bias towards the null. While some metabolites like MEP and MBP are relatively stable over time, others show greater variability (Braun et al., 2012). In addition, we unexpectedly observed that median concentrations of all phthalate metabolites decreased from Visit 1 to Visit 2. While it is possible that this difference could reflect pharmacokinetic changes across pregnancy, other recent studies (including several with phthalate analyses conducted at the same lab as in this analysis) have not reported systematic decreases in phthalate metabolites across pregnancy. In the GAPPS cohort, 10 of 16 metabolites measured decreased from the second to the third trimester (Paquette et al., 2021), while in Generation R, only 4 of 11 metabolites decreased (Santos et al., 2021), and in the PROGRESS study, nearly all phthalate metabolite concentrations non-significantly increased over that period (Wu et al., 2020). We considered laboratory batch effects as a possible explanation, however extensive quality control and assurance (QA/QC) procedures were conducted at the time of assay (and revisited at the time of this analysis); despite the unexpected difference in median values, no QA/QC concerns were noted, nor were there any systematic differences in the way the samples were collected and processed across visits. An additional complication resulting from the difference in phthalate metabolite concentrations across the two visits is that it confounds our ability to evaluate the possibility that phthalates exert non-monotonic effects on CRH; that is, the impacts of phthalates on CRH may differ at high and low concentrations, a possibility suggested by some prior work on endocrine disruptors (Zoeller and Vandenberg, 2015). That said, our within-visit analyses indicated predominantly linear associations, with the exception of a small set of metabolites for which slopes changed at the highest concentrations ($\geq 95^{\text{th}}$ percentile). In light of the unexpected and unexplained decrease in metabolite concentrations across pregnancy, additional work to replicate these findings will be important.

An additional limitation is that we measured pCRH at only two timepoints, limiting our ability to characterize the rise with more sophisticated modelling approaches. While the CANDLE cohort was representative of Shelby County, TN, our results may not be generalizable to the U.S. population as a whole. In addition, while our findings on phthalates and pregnancy complications are thought-provoking, the study was not specifically designed to examine pregnancy complications, and in fact excluded women with several major medical issues at enrollment, thus the numbers of women who developed gestational diabetes and gestational hypertension were relatively small (59 and 102, respectively). That said, our study provides insight into environmental exposures, pCRH, and pregnancy complications among women who were healthy at baseline. Future work focused on women with these

complications is needed to examine the extent to which they may be particularly susceptible to endocrine disruption. Finally, given the large number of metabolites considered in the secondary individual linear regression models, results from those models should be interpreted with caution.

In summary, in this large, diverse U.S. cohort, associations between phthalate concentrations and pCRH varied across gestation and in relation to pregnancy complications. Alterations in pCRH concentrations in pregnancy may have profound implications for the course of pregnancy as well as subsequent child health and development. In addition to risks related to fetal growth restriction and preterm birth (Wadhwa et al., 2004; Lee, 2014), gestational pCRH exposures may impact maternal postpartum psychiatric health as well as infant and child biology, temperament, behavior (Davis et al., 2005; Sandman et al., 2018; Yim et al., 2009; Glynn and Sandman, 2014; Howland et al., 2016). With ours being the largest study on this topic to date, more work is needed to examine the extent to which pCRH may be vulnerable to dysregulation by phthalates as well as the many endocrine disrupting chemicals in the modern environment and to explore the possibility that women with pregnancy complications may have heightened vulnerability to endocrine disruptors.

Authors' roles

EB conceptualized the current analysis and led manuscript writing; MC, ST, and DD led data analysis and interpretation; NB, FT, KL, AS, SS, CL, and CK were involved in obtaining funding for this work as well as study design and implementation; KK and RS led biomarker analyses; all authors edited the manuscript and approved of the final version.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We thank the CANDLE participants as well as the staff of CANDLE and ECHO PATHWAYS.

Study funding/competing interest(s)

The ECHO PATHWAYS Consortium is funded by NIH UG3OD023271 and UH3OD023271. The CANDLE study is funded by the Urban Child Institute as well as CIHR award number MWG-146331. Additional support for this analysis was provided by NIH P30ES005022, P30ES001247, and T32ES007271.

Data availability

The data utilized for this study are not publicly available but de-identified data may be available on request, subject to approval by the internal review board and under a formal data use agreement. Contact the corresponding author for more information.

Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.envint.2022.107078>.

References

Aker, A.M., Ferguson, K.K., Rosario, Z.Y., Mukherjee, B., Alshawabkeh, A.N., Calafat, A. M., Cordero, J.F., Meeker, J.D., 2019. A repeated measures study of phenol, paraben and Triclocarban urinary biomarkers and circulating maternal hormones during gestation in the Puerto Rico PROTECT cohort. *Environ Health*. 18 (1), 28.

Arbuckle, T.E., Davis, K., Marro, L., Fisher, M., Legrand, M., LeBlanc, A., Gaudreau, E., Foster, W.G., Choerung, V., Fraser, W.D., 2014. Phthalate and bisphenol A exposure among pregnant women in Canada—results from the MIREC study. *Environ. Int.* 68, 55–65.

Bellavia, A., Hauser, R., Seely, E.W., Meeker, J.D., Ferguson, K.K., McElrath, T.F., James-Todd, T., 2017. Urinary phthalate metabolite concentrations and maternal weight during early pregnancy. *Int. J. Hyg. Environ. Health* 220 (8), 1347–1355.

Bloom, M.S., Wenzel, A.G., Brock, J.W., Kucklick, J.R., Wineland, R.J., Cruze, L., Unal, E. R., Yucel, R.M., Jiyessova, A., Newman, R.B., 2019. Racial disparity in maternal phthalates exposure; Association with racial disparity in fetal growth and birth outcomes. *Environ. Int.* 127, 473–486.

Boeniger, M.F., Lowry, L.K., Rosenberg, J., 1993. Interpretation of urine results used to assess chemical exposure with emphasis on creatinine adjustments: a review. *Am. Ind. Hyg. Assoc. J.* 54 (10), 615–627.

Braun, J.M., Smith, K.W., Williams, P.L., Calafat, A.M., Berry, K., Ehrlich, S., Hauser, R., 2012. Variability of urinary phthalate metabolite and bisphenol A concentrations before and during pregnancy. *Environ. Health Perspect.* 120 (5), 739–745.

Buckley, J.P., Kim, H., Wong, E., Rebholz, C.M., 2019. Ultra-processed food consumption and exposure to phthalates and bisphenols in the US National Health and Nutrition Examination Survey, 2013–2014. *Environ. Int.* 131, 105057. <https://doi.org/10.1016/j.envint.2019.105057>.

Campbell, E.A., Linton, E.A., Wolfe, C.D.A., Scraggs, P.R., Jones, M.T., Lowry, P.J., 1987. Plasma corticotropin-releasing hormone concentrations during pregnancy and parturition. *J. Clin. Endocrinol. Metab.* 64 (5), 1054–1059.

Cantonwine, D.E., Meeker, J.D., Ferguson, K.K., Mukherjee, B., Hauser, R., McElrath, T. F., 2016. Urinary Concentrations of Bisphenol A and Phthalate Metabolites Measured during Pregnancy and Risk of Preeclampsia. *Environ. Health Perspect.* 124 (10), 1651–1655.

Carrico, C., Gennings, C., Wheeler, D.C., Factor-Litvak, P., 2015. Characterization of Weighted Quantile Sum Regression for Highly Correlated Data in a Risk Analysis Setting. *J. Agric. Biol. Environ. Stat.* 20 (1), 100–120.

Cathey, A.L., Watkins, D., Rosario, Z.Y., Vélez, C., Alshawabkeh, A.N., Cordero, J.F., Meeker, J.D., 2019. Associations of Phthalates and Phthalate Replacements With CRH and Other Hormones Among Pregnant Women in Puerto Rico. *J. Endocr. Soc.* 3 (6), 1127–1149.

Chan, M., Mita, C., Bellavia, A., Parker, M., James-Todd, T., 2021. Racial/Ethnic Disparities in Pregnancy and Prenatal Exposure to Endocrine-Disrupting Chemicals Commonly Used in Personal Care Products. *Curr. Environ. Health Rep.* 8 (2), 98–112.

Clifton, V.L., Read, M.A., Leitch, I.M., Giles, W.B., Boura, A.L., Robinson, P.J., Smith, R., 1995. Corticotropin-releasing hormone-induced vasodilatation in the human fetal-placental circulation: involvement of the nitric oxide-cyclic guanosine 3',5'-monophosphate-mediated pathway. *J. Clin. Endocrinol. Metab.* 80 (10), 2888–2893.

Curtin, P., Kellogg, J., Cech, N., Gennings, C., 2021. A random subset implementation of weighted quantile sum (WQSR) regression for analysis of high dimensional mixtures. *Commun. Stat. – Simulat. Comput.* 50 (4), 1119–1134.

Davis, E.P., Glynn, L.M., Dunkel Schetter, C., Hobel, C., Chicé-Demet, A., Sandman, C.A., 2005. Corticotropin-releasing hormone during pregnancy is associated with infant temperament. *Dev. Neurosci.* 27 (5), 299–305.

Day, D.B., Collett, B.R., Barrett, E.S., Bush, N.R., Swan, S.H., Nguyen, R.H.N., Szpiro, A. A., Sathyanarayana, S., 2021. Phthalate mixtures in pregnancy, autistic traits, and adverse childhood behavioral outcomes. *Environ. Int.* 147, 106330. <https://doi.org/10.1016/j.envint.2020.106330>.

Dorman, D.C., Chiu, W., Hales, B.F., Hauser, R., Johnson, K.J., Mantus, E., Martel, S., Robinson, K.A., Rooney, A.A., Rudel, R., Sathyanarayana, S., Schantz, S.L., Waters, K.M., 2018. Systematic reviews and meta-analyses of human and animal evidence of prenatal diethylhexyl phthalate exposure and changes in male anogenital distance. *J. Toxicol. Environ. Health B Crit. Rev.* 21 (4), 207–226.

Eick, S.M., Goin, D.E., Cushing, L., DeMico, E., Smith, S., Park, J.S., Padula, A.M., Woodruff, T.J., Morello-Frosch, R., 2021. Joint effects of prenatal exposure to per- and poly-fluoroalkyl substances and psychosocial stressors on corticotropin-releasing hormone during pregnancy. *J. Expo Sci. Environ. Epidemiol.*

Ferguson, K.K., Chin, H.B., 2017. Environmental chemicals and preterm birth: Biological mechanisms and the state of the science. *Curr. Epidemiol. Rep.* 4 (1), 56–71.

Ferguson, K.K., McElrath, T.F., Meeker, J.D., 2014. Environmental phthalate exposure and preterm birth. *JAMA Pediatr.* 168 (1), 61–67.

Ferguson, K.K., McElrath, T.F., Ko, Y.-A., Mukherjee, B., Meeker, J.D., 2014. Variability in urinary phthalate metabolite levels across pregnancy and sensitive windows of exposure for the risk of preterm birth. *Environ. Int.* 70, 118–124.

Ferguson, K.K., Rosen, E.M., Rosario, Z., Feric, Z., Calafat, A.M., McElrath, T.F., Vélez Vega, C., Cordero, J.F., Alshawabkeh, A., Meeker, J.D., 2019. Environmental phthalate exposure and preterm birth in the PROTECT birth cohort. *Environ. Int.* 132, 105099. <https://doi.org/10.1016/j.envint.2019.105099>.

Ferguson, K.K., Rosen, E.M., Barrett, E.S., Nguyen, R.H.N., Bush, N., McElrath, T.F., Swan, S.H., Sathyanarayana, S., 2019. Joint impact of phthalate exposure and stressful life events in pregnancy on preterm birth. *Environ. Int.* 133, 105254. <https://doi.org/10.1016/j.envint.2019.105254>.

Fisher, B.G., Frederiksen, H., Andersson, A.M., Juul, A., Thankamony, A., Ong, K.K., Dunger, D.B., Hughes, I.A., Acerini, C.L., 2018. Serum Phthalate and Triclosan Levels Have Opposing Associations With Risk Factors for Gestational Diabetes Mellitus. *Front. Endocrinol. (Lausanne)*. 9, 99.

Frim, D.M., Emanuel, R.L., Robinson, B.G., Smas, C.M., Adler, G.K., Majzoub, J.A., 1988. Characterization and gestational regulation of corticotropin-releasing hormone messenger RNA in human placenta. *J. Clin. Invest.* 82 (1), 287–292.

- Gangestad, S.W., Caldwell Hooper, A.E., Eaton, M.A., 2012. On the function of placental corticotropin-releasing hormone: a role in maternal-fetal conflicts over blood glucose concentrations. *Biol. Rev. Camb. Philos. Soc.* 87 (4), 856–873.
- Gao, L., Lv, C., Xu, C., Li, Y., Cui, X., Gu, H., Ni, X., 2012. Differential regulation of glucose transporters mediated by CRH receptor type 1 and type 2 in human placental trophoblasts. *Endocrinology* 153 (3), 1464–1471.
- Gao, H., Wang, Y.-F., Huang, K., Han, Y., Zhu, Y.-D., Zhang, Q.-F., Xiang, H.-Y., Qi, J., Feng, L.-L., Zhu, P., Hao, J.-h., Tao, X.-G., Tao, F.-B., 2019. Prenatal phthalate exposure in relation to gestational age and preterm birth in a prospective cohort study. *Environ. Res.* 176, 108530. <https://doi.org/10.1016/j.envres.2019.108530>.
- Glynn, L.M., Sandman, C.A., 2014. Evaluation of the association between placental corticotropin-releasing hormone and postpartum depressive symptoms. *Psychosom. Med.* 76 (5), 355–362.
- Gore, A.C., Chappell, V.A., Fenton, S.E., Flaws, J.A., Nadal, A., Prins, G.S., Toppari, J., Zoeller, R.T., 2015. EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. *Endocr. Rev.* 36 (6), E1–E150.
- Guo, Y., Weck, J., Sundaram, R., Goldstone, A.E., Buck Louis, G., Kannan, K., 2014. Urinary concentrations of phthalates in couples planning pregnancy and its association with 8-hydroxy-2'-deoxyguanosine, a biomarker of oxidative stress: longitudinal investigation of fertility and the environment study. *Environ. Sci. Technol.* 48 (16), 9804–9811.
- Harley, K.G., Berger, K.P., Kogut, K., Parra, K., Lustig, R.H., Greenspan, L.C., Calafat, A.M., Ye, X., Eskenazi, B., 2019. Association of phthalates, parabens and phenols found in personal care products with pubertal timing in girls and boys. *Hum. Reprod.* 34 (1), 109–117.
- Harville, E.W., Savitz, D.A., Dole, N., Herring, A.H., Thorp, J.M., Light, K.C., 2008. Stress and placental resistance measured by Doppler ultrasound in early and mid-pregnancy. *Ultrasound. Obstet. Gynecol.* 32 (1), 23–30.
- Harville, E.W., Savitz, D.A., Dole, N., Herring, A.H., Thorp, J.M., 2009. Stress questionnaires and stress biomarkers during pregnancy. *J. Womens Health (Larchmt)*. 18 (9), 1425–1433.
- Herrera, C.L., Bowman, M.E., McIntire, D.D., Nelson, D.B., Smith, R., 2021. Revisiting the placental clock: Early corticotropin-releasing hormone rise in recurrent preterm birth. *PLoS One*. 16 (9) e0257422.
- Hobel, C.J., Dunkel-Schetter, C., Roesch, S.C., Castro, L.C., Arora, C.P., 1999. Maternal plasma corticotropin-releasing hormone associated with stress at 20 weeks' gestation in pregnancies ending in preterm delivery. *Am. J. Obstet. Gynecol.* 180 (1), S257–S263.
- Hogg, K., Blair, J.D., McFadden, D.E., von Dadelszen, P., Robinson, W.P., 2013. Early onset pre-eclampsia is associated with altered DNA methylation of cortisol-signalling and steroidogenic genes in the placenta. *PLoS One*. 8 (5), e62969.
- Hornung, R.W., Reed, L.D., 1990. Estimation of Average Concentration in the Presence of Nondetectable Values. *Appl. Occup. Environ. Hyg.* 5 (1), 46–51.
- Howland, M.A., Sandman, C.A., Glynn, L.M., Crippen, C., Davis, E.P., 2016. Fetal exposure to placental corticotropin-releasing hormone is associated with child self-reported internalizing symptoms. *Psychoneuroendocrinology*. 67, 10–17.
- Hu, J.M.Y., Arbuckle, T.E., Janssen, P., Lanphear, B.P., Braun, J.M., Platt, R.W., Chen, A., Fraser, W.D., McCandless, L.C., 2020. Associations of prenatal urinary phthalate exposure with preterm birth: The Maternal-Infant Research on Environmental Chemicals (MIREC) Study. *Can. J. Public Health* 111 (3), 333–341.
- James-Todd, T.M., Meeker, J.D., Huang, T., Hauser, R., Ferguson, K.K., Rich-Edwards, J.W., McElrath, T.F., Seely, E.W., 2016. Pregnancy urinary phthalate metabolite concentrations and gestational diabetes risk factors. *Environ. Int.* 96, 118–126.
- James-Todd, T.M., Chiu, Y.-H., Zota, A.R., 2016. Racial/ethnic disparities in environmental endocrine disrupting chemicals and women's reproductive health outcomes: epidemiological examples across the life course. *Curr. Epidemiol. Rep.* 3 (2), 161–180.
- Kannan, K., Stathis, A., Mazzella, M.J., Andra, S.S., Barr, D.B., Hecht, S.S., Merrill, L.S., Galusha, A.L., Parsons, P.J., 2021. Quality assurance and harmonization for targeted biomonitoring measurements of environmental organic chemicals across the Children's Health Exposure Analysis Resource laboratory network. *Int. J. Hyg. Environ. Health* 234, 113741. <https://doi.org/10.1016/j.ijheh.2021.113741>.
- Karteris, E., Goumenou, A., Koumantakis, E., Hillhouse, E.W., Grammatopoulos, D.K., 2003. Reduced expression of corticotropin-releasing hormone receptor type-1 alpha in human preeclamptic and growth-restricted placentas. *J. Clin. Endocrinol. Metab.* 88 (1), 363–370.
- Karteris, E., Vatish, M., Hillhouse, E.W., Grammatopoulos, D.K., 2005. Preeclampsia is associated with impaired regulation of the placental nitric oxide-cyclic guanosine monophosphate pathway by corticotropin-releasing hormone (CRH) and CRH-related peptides. *J. Clin. Endocrinol. Metab.* 90 (6), 3680–3687.
- Koch, H.M., Preuss, R., Angerer, J., 2006. Di(2-ethylhexyl)phthalate (DEHP): human metabolism and internal exposure— an update and latest results. *Int. J. Androl.* 29 (1), 155–165. ; discussion 181–155.
- Kubany, E.S., Leisen, M.B., Kaplan, A.S., Kelly, M.P., 2000. Validation of a brief measure of posttraumatic stress disorder: the Distressing Event Questionnaire (DEQ). *Psychol. Assess.* 12 (2), 197–209.
- Laatikainen, T.J., 1991. Corticotropin-releasing hormone and opioid peptides in reproduction and stress. *Ann. Med.* 23 (5), 489–496.
- Laatikainen, T., Virtanen, T., Kaaja, R., Salminen-Lappalainen, K., 1991. Corticotropin-releasing hormone in maternal and cord plasma in pre-eclampsia. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 39 (1), 19–24.
- Latendresse, G., Ruiz, R.J., 2008. Bioassay research methodology: measuring CRH in pregnancy. *Biol. Res. Nurs.* 10 (1), 54–62.
- Lee, C., 2014. Intergenerational health consequences of in utero exposure to maternal stress: evidence from the 1980 Kwangju uprising. *Soc. Sci. Med.* 119, 284–291.
- Liapi, C.A., Tsakalia, D.E., Panitsa-Fafila, C.C., Antsaklis, A.I., Aravantinos, D.I., Batrinos, M.L., 1996. Corticotropin-releasing-hormone levels in pregnancy-induced hypertension. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 68 (1–2), 109–114.
- Makriganakis, A., Semmler, M., Briesse, V., Eckerle, H., Minas, V., Mylonas, I., Friese, K., Jeschke, U., 2007. Maternal serum corticotropin-releasing hormone and ACTH levels as predictive markers of premature labor. *Int. J. Gynaecol. Obstet.* 97 (2), 115–119.
- Makriganakis, A., Vrekoussis, T., Zoumakis, E., Navrozoglou, I., Kalantaridou, S.N., 2018. CRH Receptors in Human Reproduction. *Curr. Mol. Pharmacol.* 11 (1), 81–87.
- Martínez Steele, E., Khandpur, N., da Costa Louzada, M., Téllez-Rojo, C.A., 2020. Association between dietary contribution of ultra-processed foods and urinary concentrations of phthalates and bisphenol in a nationally representative sample of the US population aged 6 years and older. *PLoS One*. 15 (7), e0236738.
- Martínez-Razo, L.D., Martínez-Ibarra, A., Vázquez-Martínez, E.R., Cerbón, M., 2021. The impact of Di-(2-ethylhexyl) Phthalate and Mono(2-ethylhexyl) Phthalate in placental development, function, and pathophysiology. *Environ. Int.* 146, 106228. <https://doi.org/10.1016/j.envint.2020.106228>.
- Meeker, J.D., Hu, H., Cantonwine, D.E., Lamadrid-Figueroa, H., Calafat, A.M., Ettinger, A.S., Hernandez-Avila, M., Loch-Carusio, R., Téllez-Rojo, M.M., 2009. Urinary phthalate metabolites in relation to preterm birth in Mexico city. *Environ. Health Perspect.* 117 (10), 1587–1592.
- Moog, N.K., Buss, C., Entringer, S., Shahbaba, B., Gillen, D.L., Hobel, C.J., Wadhwa, P.D., 2016. Maternal Exposure to Childhood Trauma Is Associated During Pregnancy With Placental-Fetal Stress Physiology. *Biol. Psychiatry* 79 (10), 831–839.
- Paquette, A.G., MacDonald, J., Lapehn, S., Bammiller, T., Kruger, L., Day, D.B., Price, N.D., Lofcus, C., Kannan, K., Marsit, C., Mason, W.A., Bush, N.R., LeWinn, K.Z., Enquobahrie, D.A., Prasad, B., Karr, C.J., Sathyanarayana, S., 2021. A Comprehensive Assessment of Associations between Prenatal Phthalate Exposure and the Placental Transcriptomic Landscape. *Environ. Health Perspect.* 129 (9), 097003. <https://doi.org/10.1289/EHP9703>.
- Perkins, A.V., Eben, F., Wolfe, C.D., Schulte, H.M., Linton, E.A., 1993. Plasma measurements of corticotropin-releasing hormone-binding protein in normal and abnormal human pregnancy. *J. Endocrinol.* 138 (1), 149–157.
- Perkins, A.V., Linton, E.A., Eben, F., Simpson, J., Wolfe, C.D.A., Redman, C.W.G., 1995. Corticotropin-releasing hormone and corticotropin-releasing hormone binding protein in normal and pre-eclamptic human pregnancies. *Br. J. Obstet. Gynaecol.* 102 (2), 118–122.
- Phillips, E.M., Trasande, L., Kahn, L.G., Gaillard, R., Steegers, E.A.P., Jaddoe, V.W.V., 2019. Early pregnancy bisphenol and phthalate metabolite levels, maternal hemodynamics and gestational hypertensive disorders. *Hum. Reprod.* 34 (2), 365–373.
- Purwosunu, Y., Sekizawa, A., Farina, A., Wibowo, N., Okazaki, S., Nakamura, M., Samura, O., Fujito, N., Okai, T., 2007. Cell-free mRNA concentrations of CRH, PLAC1, and selectin-P are increased in the plasma of pregnant women with preeclampsia. *Prenat. Diagn.* 27 (8), 772–777.
- Renzetti, S., Curtin, P., Just, A., Bello, G., Gennings, C., 2021. gWQS. R package version 3.0.4.
- Rocha, B.A., Asimakopoulos, A.G., Barbosa, F., Kannan, K., 2017. Urinary concentrations of 25 phthalate metabolites in Brazilian children and their association with oxidative DNA damage. *Sci. Total Environ.* 586, 152–162.
- Ruiz, R.J., Gennaro, S., O'Connor, C., Dwivedi, A., Gibeau, A., Keshinover, T., Welsh, T., 2016. CRH as a Predictor of Preterm Birth in Minority Women. *Biol. Res. Nurs.* 18 (3), 316–321.
- Sandman, C.A., 2018. Prenatal CRH: An integrating signal of fetal distress. *Dev. Psychopathol.* 30 (3), 941–952.
- Sandman, C.A., Glynn, L., Schetter, C.D., Wadhwa, P., Garite, T., Chicx-DeMet, A., Hobel, C., 2006. Elevated maternal cortisol early in pregnancy predicts third trimester levels of placental corticotropin releasing hormone (CRH): priming the placental clock. *Peptides* 27 (6), 1457–1463.
- Sandman, C.A., Curran, M.M., Davis, E.P., Glynn, L.M., Head, K., Baram, T.Z., 2018. Cortical Thinning and Neuropsychiatric Outcomes in Children Exposed to Prenatal Adversity: A Role for Placental CRH? *Am. J. Psychiatry* 175 (5), 471–479.
- Santos, S., Sol, C.M., van Zwol – Janssens, C., Phillips, E.M., Asimakopoulos, A.G., Martínez-Moral, M.-P., Kannan, K., Jaddoe, V.W.V., Trasande, L., 2021. Maternal phthalate urine concentrations, fetal growth and adverse birth outcomes. A population-based prospective cohort study. *Environ. Int.* 151, 106443. <https://doi.org/10.1016/j.envint.2021.106443>.
- Shaffer, R.M., Ferguson, K.K., Sheppard, L., James-Todd, T., Butts, S., Chandrasekaran, S., Swan, S.H., Barrett, E.S., Nguyen, R., Bush, N., McElrath, T.F., Sathyanarayana, S., 2019. Maternal urinary phthalate metabolites in relation to gestational diabetes and glucose intolerance during pregnancy. *Environ. Int.* 123, 588–596.
- Shoaff, J., Papandonatos, G.D., Calafat, A.M., Ye, X., Chen, A., Lanphear, B.P., Yolton, K., Braun, J.M., 2017. Early-Life Phthalate Exposure and Adiposity at 8 Years of Age. *Environ. Health Perspect.* 125 (9), 097008. <https://doi.org/10.1289/EHP1022>.
- Simpson, S.J.S., Smith, L.L.F., Jones, P.M., Bowe, J.E., 2020. UCN2: a new candidate influencing pancreatic β -cell adaptations in pregnancy. *J. Endocrinol.* 245 (2), 247–257.
- Sloven, N., Roberts, A.L., LeWinn, K.Z., Bush, N.R., Rovnaghi, C.R., Tylavsky, F., Anand, K.J.S., 2018. Maternal experiences of trauma and hair cortisol in early childhood in a prospective cohort. *Psychoneuroendocrinology*. 98, 168–176.
- Smith, R., Mesiano, S., McGrath, S., 2002. Hormone trajectories leading to human birth. *Regul. Pept.* 108 (2–3), 159–164.
- Smith, R., Smith, J.I., Shen, X., Engel, P.J., Bowman, M.E., McGrath, S.A., Bisits, A.M., McElduff, P., Giles, W.B., Smith, D.W., 2009. Patterns of plasma corticotropin-

- releasing hormone, progesterone, estradiol, and estriol change and the onset of human labor. *J. Clin. Endocrinol. Metab.* 94 (6), 2066–2074.
- Sontag-Padilla, L., Burnms, R., Shih, R., Griffin, B., Martin, L., Chandra, A., Tylavsky, F., 2015. The Urban Child Institute CANDLER Study: Methodological Overview and Baseline Sample description. RAND Corporation.
- Steine, I.M., LeWinn, K.Z., Lisha, N., Tylavsky, F., Smith, R., Bowman, M., Sathyanarayana, S., Karr, C.J., Smith, A.K., Kobor, M., Bush, N.R., 2020. Maternal exposure to childhood traumatic events, but not multi-domain psychosocial stressors, predict placental corticotropin releasing hormone across pregnancy. *Soc. Sci. Med.* 266, 113461. <https://doi.org/10.1016/j.socscimed.2020.113461>.
- Sunderland, E.M., Hu, X.C., Dassuncao, C., Tokranov, A.K., Wagner, C.C., Allen, J.G., 2019. A review of the pathways of human exposure to poly- and perfluoroalkyl substances (PFASs) and present understanding of health effects. *J. Expo Sci. Environ. Epidemiol.* 29 (2), 131–147.
- Swan, S.H., Sathyanarayana, S., Barrett, E.S., Janssen, S., Liu, F., Nguyen, R.H.N., Redmon, J.B., the TIDES Study Team, Liu, F., Scher, E., Stasenko, M., Ayash, E., Schirmer, M., Farrell, J., Thiet, M.-P., Baskin, L., Gray Chelsea Georgesen, H.L., Rody, B.J., Terrell, C.A., Kaur, K., Brantley, E., Fiore, H., Kochman, L., Parlett, L., Marino, J., Hulbert, W., Mevorach, R., Pressman, E., Ivicsek, K., Salveson, B., Alcedo, G., 2015. First trimester phthalate exposure and anogenital distance in newborns. *Hum. Reprod.* 30 (4), 963–972.
- Taylor, A.L., Fishman, L.M., 1988. Corticotropin-releasing hormone. *N. Engl. J. Med.* 319 (4), 213–222.
- Thomson, M., 2013. The physiological roles of placental corticotropin releasing hormone in pregnancy and childbirth. *J. Physiol. Biochem.* 69 (3), 559–573.
- Tyson, E.K., Smith, R., Read, M., 2009. Evidence that corticotropin-releasing hormone modulates myometrial contractility during human pregnancy. *Endocrinology* 150 (12), 5617–5625.
- Vale, W., Spiess, J., Rivier, C., Rivier, J., 1981. Characterization of a 41-residue ovine hypothalamic peptide that stimulates secretion of corticotropin and beta-endorphin. *Science* 213 (4514), 1394–1397.
- Valsamakis, G., Papatheodorou, D., Chalarakis, N., Manolikaki, M., Margeli, A., Papassotiriou, I., Barber, T.M., Kumar, S., Kalantaridou, S., Mastorakos, G., 2020. Maternal chronic stress correlates with serum levels of cortisol, glucose and C-peptide in the fetus, and maternal non chronic stress with fetal growth. *Psychoneuroendocrinology*. 114, 104591. <https://doi.org/10.1016/j.psyneuen.2020.104591>.
- Wadhwa, P.D., Garite, T.J., Porto, M., Glynn, L., Chicx-DeMet, A., Dunkel-Schetter, C., Sandman, C.A., 2004. Placental corticotropin-releasing hormone (CRH), spontaneous preterm birth, and fetal growth restriction: a prospective investigation. *Am. J. Obstet. Gynecol.* 191 (4), 1063–1069.
- Wang, X.K., Agarwal, M., Parobchak, N., Rosen, A., Vetrano, A.M., Srinivasan, A., Wang, B., Rosen, T., 2016. Mono-(2-Ethylhexyl) Phthalate Promotes Pro-Labor Gene Expression in the Human Placenta. *PLoS One*. 11 (1), e0147013.
- Wang, Y., Zhu, H., Kannan, K., 2019. A Review of Biomonitoring of Phthalate Exposures. *Toxics*. 7 (2), 21. <https://doi.org/10.3390/toxics7020021>.
- Warembourg, C., Basagaña, X., Seminati, C., de Bont, J., Granum, B., Lyon-Caen, S., Manzano-Salgado, C.B., Pin, I., Sakhi, A.K., Siroux, V., Slama, R., Urquiza, J., Vrijheid, M., Thomsen, C., Casas, M., 2019. Exposure to phthalate metabolites, phenols and organophosphate pesticide metabolites and blood pressure during pregnancy. *Int. J. Hyg. Environ. Health* 222 (3), 446–454.
- Warner, G.R., Dettogni, R.S., Bagchi, I.C., Flaws, J.A., Graceli, J.B., 2021. Placental outcomes of phthalate exposure. *Reprod. Toxicol.* 103, 1–17.
- Werner, E.F., Braun, J.M., Yolton, K., Khoury, J.C., Lanphear, B.P., 2015. The association between maternal urinary phthalate concentrations and blood pressure in pregnancy: The HOME Study. *Environ. Health*. 14, 75.
- White, H., 1980. A heteroskedasticity-consistent covariance matrix estimator and a direct test for heteroskedasticity. *Econometrica*. 48, 817–838.
- Woodruff, T.J., Zota, A.R., Schwartz, J.M., 2011. Environmental chemicals in pregnant women in the United States: NHANES 2003–2004. *Environ. Health Perspect.* 119 (6), 878–885.
- Wu, H., Kupsco, A.J., Deierlein, A.L., Just, A.C., Calafat, A.M., Oken, E., Braun, J.M., Mercado-Garcia, A., Cantoral, A., Téllez-Rojo, M.M., Wright, R.O., Baccarelli, A.A., 2020. Trends and Patterns of Phthalates and Phthalate Alternatives Exposure in Pregnant Women from Mexico City during 2007–2010. *Environ. Sci. Technol.* 54 (3), 1740–1749.
- Yim, I.S., Glynn, L.M., Dunkel-Schetter, C., Hobel, C.J., Chicx-DeMet, A., Sandman, C.A., 2009. Risk of postpartum depressive symptoms with elevated corticotropin-releasing hormone in human pregnancy. *Arch. Gen. Psychiatry* 66 (2), 162–169.
- Zhong, Q.i., Liu, H.-l., Fu, H., Niu, Q.-S., Wu, H.-B., Huang, F., 2021. Prenatal exposure to phthalates with preterm birth and gestational age: A systematic review and meta-analysis. *Chemosphere* 282, 130991. <https://doi.org/10.1016/j.chemosphere.2021.130991>.
- Zoeller, R.T., Vandenberg, L.N., 2015. Assessing dose-response relationships for endocrine disrupting chemicals (EDCs): a focus on non-monotonicity. *Environ. Health*. 14, 42.
- Zota, A.R., Phillips, C.A., Mitro, S.D., 2016. Recent Fast Food Consumption and Bisphenol A and Phthalates Exposures among the U.S. Population in NHANES, 2003–2010. *Environ. Health Perspect.* 124 (10), 1521–1528.
- Zota, A.R., Shamasunder, B., 2017. The environmental injustice of beauty: framing chemical exposures from beauty products as a health disparities concern. *Am. J. Obstet. Gynecol.* 217 (4) e411–418.e416.
- Zukin, H., Eskenazi, B., Holland, N., Harley, K.G., 2021. Prenatal exposure to phthalates and maternal metabolic outcomes in a high-risk pregnant Latina population. *Environ. Res.* 194, 110712. <https://doi.org/10.1016/j.envres.2021.110712>.